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## Hepatic encephalopathy in dogs

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Malfunctioning of the neutralizing function of the liver leads to the buildup of toxins in the body, which negatively impacts the central nervous system. The objective of our study was examining dogs that had typical concurrent symptoms of dysfunctions of the liver and central nervous system. We studied two groups of dogs – the control (clinically healthy) and experimental (sick dogs with symptoms of disorders of the liver and brain). Each of the groups consisted of 10 animals: four Yorkshire Terriers, two Maltese dogs, two Russian Toy dogs, one English Cocker Spaniel, and one mixed-breed dog. The animals were examined clinically, underwent ultrasound diagnostics, and had their blood collected for laboratory analyses. Clinically, in all the sick dogs, we identified the typical symptoms of liver lesions – pain in the liver region, increase in its area, and also malfunctioning of the central nervous system manifesting in soporous state, ataxia, and spasms. In blood plasma of all the sick dogs, we observed significant increase in the concentration of ammonia. High ammonia content in blood of dogs indicates an impaired neutralizing function of the liver. At the same time, it is the main endotoxin that affects the central nervous system, promoting the development of liver encephalopathy. The level of hyperammonemia was closely associated with the severity of the course of pathology. In blood serum of the dogs with signs of liver encephalopathy, the content of bile acids increased 4.5-fold and total bilirubin increased 31%, while albumin decreased 15%. In the blood of 60% of the patients, the level of  $\text{Na}^+$  was low. In serum of blood from the patients, the activities of AST, ALT, and AP were increased. The ultrasound studies of the liver revealed increased echotexture and impaired hemodynamics, which, when co-occurring with high activity of indicator enzymes, indicate damage to liver cells. During severe liver encephalopathy, the dogs were found to have leukocytosis, neutrophilia, and lymphocytopenia, and also decline in the content of hemoglobin, number of erythrocytes, and hematocrit value. The next stage of our research will focus on the role of portosystemic shunts in the development of hepatic encephalopathy in dogs.

**Keywords:** dogs; liver lesion; brain lesion; ammonia; metabolism; ultrasound diagnostics.

### Introduction

Liver diseases of various etiologies are often observed in dogs, depending on their age, sex, and breed (Rothuizen, 2009; Assawarachan et al., 2023; Kashliak & Vlizlo, 2023). During pathology of the liver, its main functions are impaired, in particular, neutralizing exo- and endotoxins. Hepatic cells take part in detoxification of bacterial toxins, mycotoxins, phytotoxins, various toxins of organic and synthetic origins, and also endotoxins formed during breakdown of food in the gastrointestinal tract (Besa et al., 2012). Only a minimum amount of toxins enters the general blood circulation, because the neutralizing function of the liver is quite stable and even during a liver failure can be sustained at the physiological level (Gow, 2017). Neutralization of exo- and endotoxins in the liver malfunctions in cases of significant damage to hepatocytes or the development of shunting. At the same time, toxins infiltrate the blood and concentrate in the bodies of sick animals, thus involving the central nervous system in the pathological process (Wang et al., 2017; Kabaria et al., 2021; Lynch, 2023). Such sick animals are diagnosed with lesions of the liver and brain. This pathology was described as hepatic encephalopathy (hepatogenic encephalopathy, hepatocerebral syndrome) (Gluud et al., 2016; Hadjihambi et al., 2019).

Depending on etiology, in dogs and humans, there are three types of development of hepatic encephalopathy: type A occurs during acute liver failure; type B is related to congenital portosystemic shunts; type C develops in cases of liver cirrhosis and portal hypertension, when acquired portosystemic shunts form (Kraun et al., 2014; Lidbury et al., 2015; Swaminathan et al., 2018). During the pathogenesis of hepatic encephalopathy, the key role is believed to be played by ammonia (Vlizlo, 1999; Ti-

vers et al., 2014; Bellafante et al., 2024). It forms in the gastrointestinal tract during catabolism of proteins. Considering that dogs are carnivores, the diet of which is based on high content of proteins, the ammonia formation in them is especially active. However, in the liver of healthy animals, it is detoxified, allowing only a small amount to circulate in blood. When the neutralizing function of hepatocytes is impaired, ammonia accumulates in the body (Caporali et al., 2015; Lidbury et al., 2016). The significance of ammonia in the development of hepatic encephalopathy is explained by the fact that it is an endotoxin, which causes pathological impact on the brain (Lima et al., 2019). In the central nervous system, ammonia leads to dysfunction of astrocytes and development of oxidative stress, leading to edema of the brain and intracranial hypertension (Romero-Gómez et al., 2015; Levitt & Levitt, 2018).

Hepatogenic encephalopathy in dogs is diagnosed in complex based on a thoroughly collected anamnesis, along with clinical, laboratory, and instrumental methods of research (Assawarachan et al., 2020). In most cases, hepatic encephalopathy in dogs is only evident after the emergence of clinical signs. Patients experience ataxia, move in circles, suffer spasms, tremor, paresis, stupor, which can lead to coma, and even death (Krishnarao & Gordon, 2020; Rose et al., 2020; Konstantinidis et al., 2023). The prognosis for such dogs is unfavorable, because the treatment efficacy is low (Mullins et al., 2022). Considering this fact, it is important to search for methods of diagnosing hepatic encephalopathy at the subclinical stage of pathology development. Important laboratory parameters in cases of this condition are the content of metabolites of protein catabolism in the blood of patients – ammonium and urea (Lawrence & Steiner, 2017). Ammonia concentration in the blood is an informative marker of hepatic encephalopathy (Tivers et al., 2014; Lima et al., 2019). Nonetheless, there

is a report that individual dogs suffering hepatocerebral syndrome had the ammonia content in blood at the level of physiological values (Gow, 2017). The objectives of the study were to examine dogs with the symptoms of lesions of the liver and central nervous system, conducting ultrasound studies of the internal organs, performing laboratory analysis of blood, and measuring the parameters that can be informative in cases of hepatic encephalopathy.

## Materials and methods

All the procedures with the animals were performed according to the European Convention for the Protection of Vertebrate Animals used for Experimental and other Scientific Purposes (Strasbourg, 1986) and the General Ethical Principles of Experiments on Animals, adopted by the First National Congress of Bioethics (Kyiv, 2001). The experiments were conducted with adherence to the principles of humanity, provided in the Guideline of the European Community (Directive 2010/63/EU, 2010).

The studies were conducted at the veterinary clinic Merlion in the city of Lviv. The material for the studies was 10 clinically healthy dogs and 10 sick ones, diagnosed with the symptoms characteristic of the hepatic encephalopathy. The clinically healthy dogs were selected in relation to the sick ones according to the analogue principle, taking into account breed and age. Each group contained four Yorkshire Terriers, two Maltese dogs, two Russian Toy dogs, one English Cocker Spaniel, and one mixed-breed dog. When an animal was admitted to the clinic, the anamnesis was gathered and clinical examination was performed, and also blood samples for laboratory analyses were collected. On a hematological analyzer Mindray BC-30 Vet (Japan), we measured the content of hemoglobin, hematocrit, and the numbers of erythrocytes, leukocytes, and platelets. To generate a leukogram, we prepared and stained blood smears and counted cells under a microscope.

Using a biochemical analyzer Mindray BS-240 (Japan), we studied the blood serum for the contents of total bilirubin, total bile acids, total protein, albumin, and urea, and the activities of alanine aminotransferase (ALT), aspartate aminotransferase (AST), and alkaline phosphatase (AP). The concentration of ammonia in blood plasma was measured on a biochemical analyzer Fuji DRI-CHEM NX500 (Japan). Using an analyzer

**Table 1**

General blood analysis of the dogs ( $\bar{x} \pm SE$ ,  $n = 10$ )

Groups of animals	Hemoglobin, g/L	Erythrocytes, $10^{12}/L$	Hematocrit, L/L	Leukocytes, $10^9/L$	Eosinophile, %	Band neutrophils, %	Segmented-nucleus neutrophils	Lymphocytes, %	Monocytes, %	Platelets, $10^9/L$
Clinically healthy	171.2 $\pm$ 2.9	7.48 $\pm$ 0.24	0.45 $\pm$ 0.012	9.15 $\pm$ 0.53	2.7 $\pm$ 0.6	2.5 $\pm$ 0.6	68.2 $\pm$ 3.1	23.6 $\pm$ 2.9	3.0 $\pm$ 0.7	290 $\pm$ 25
Sick	147.6 $\pm$ 11.0*	6.71 $\pm$ 0.48	0.39 $\pm$ 0.025	15.23 $\pm$ 4.03	1.7 $\pm$ 0.4	8.0 $\pm$ 3.1	66.7 $\pm$ 3.5	19.2 $\pm$ 3.5	4.4 $\pm$ 0.8	268 $\pm$ 45

Note: \* –  $P < 0.05$  between the clinically healthy and sick animals.

In the blood plasma of the sick dogs, we found a significant increase in the ammonia concentration ( $122.6 \pm 57.2 \mu\text{mol/L}$ ;  $P < 0.001$ ), compared with the clinically healthy ones ( $23.6 \pm 9.5 \mu\text{mol/L}$ ). Hyperammonemia was observed in all the sick dogs (Fig. 1a). It has to be noted that the severity of the disease, especially nervous disorders, depended on the ammonia content in blood. When hyperammonemia was higher, hepatic encephalopathy was more acute.

The content of bile acids in blood serum of the dogs with signs of hepatic encephalopathy (Fig. 1b) measured  $56.0 \pm 35.8 \mu\text{mol/L}$ , 4.5 times higher ( $P < 0.001$ ) than in the clinically healthy dogs ( $13.1 \pm 5.4 \mu\text{mol/L}$ ).

The concentration of total bilirubin in blood (Fig. 1c) of the sick dogs was 30% higher ( $7.5 \pm 3.3 \mu\text{mol/L}$ ) than in the clinically healthy ones ( $5.7 \pm 1.5 \mu\text{mol/L}$ ). At the same time, the urea content in serum on average did not differ between the groups ( $5.93 \pm 4.4$  and  $5.90 \pm 2.1 \text{ mmol/L}$ , respectively).

In blood serum of the sick dogs, we observed decline of albumin to  $24.0 \pm 4.5 \text{ g/L}$  ( $P < 0.05$ ) compared with  $28.3 \pm 3.5 \text{ g/L}$  in the clinically healthy dogs (Fig. 1d). Hypoalbuminemia promoted decrease in the total protein in blood of the sick animals to  $65.0 \pm 7.9$ , against  $71.0 \pm 6.3 \text{ g/L}$  in the control.

The serum of the sick dogs had heightened activities of amino transferases (AST, ALT). In particular, the activity of AST in blood (Fig. 2a) was

OPTI CCA-TS2 (USA), we studied the content of electrolytes ( $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cl}^-$ ,  $\text{Ca}^{++}$ ) and pH of blood in blood serum. To perform sonography of the internal organs, we used an ultrasound apparatus Easote mylab x5 (Italy) with a 3.5–7.5 MHz sensor frequency. During the procedure, the animals were placed in the dorsal recumbency position. The sensor was placed in the region of xiphoid process and with pendulum-like movements we identified the localization and boundaries of the liver margins, the condition of parenchyma, blood vessels, bile ducts, and bladder, and also other internal organs.

The blood parameters were statistically analyzed using Statistica 7 (StatSoft Inc., USA). The graphs were developed in Statistica 7 using the generally accepted algorithms. The article presents the mean arithmetic values and standard deviation  $\bar{x} \pm SD$  (mean  $\pm$  standard deviation), presented in the figures. To compare the differences between the mean parameters of the clinically healthy and sick animals, we used the Tukey Test, where the differences were considered statistically significant at  $P < 0.05$ .

## Results

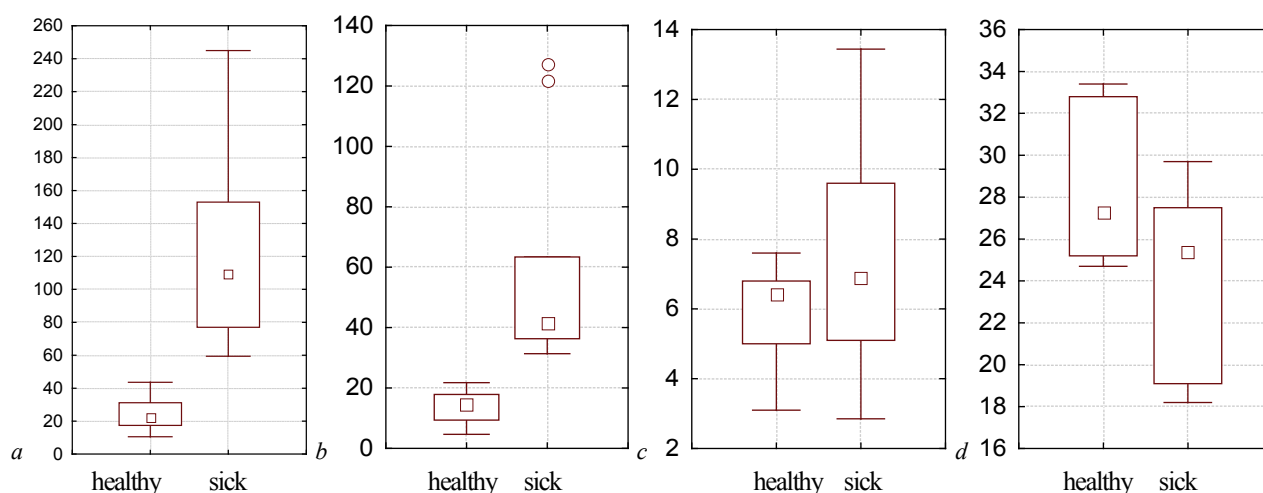
According to the owners, the dogs were depressed, had low appetite or had no appetite at all. During the clinical study of the patients, with deep palpation in the right epigastric region, we identified pain in the area of the liver and increase in its size. Of the ten patients, one was observed to have icteritiousness of the mucous membranes, and in the rest they were pale-red. At the same time, the dogs were diagnosed with the symptoms of malfunctioning of the nervous system, with inhibited general state (stupor), disorders in the coordination of moves (ataxia), and spasms. At the same time, tactile and pain sensitivity remained.

According to the results of hematological studies, the dogs with symptoms of hepatic encephalopathy had reduced level of hemoglobin ( $P < 0.05$ ). The number of erythrocytes and the size of hematocrit tended to decrease (Table 1). The mean number of platelets in blood of the dogs was no different from such in the clinically healthy dogs (Table 1) and only two out of ten had a low number of platelets. The number of leukocytes in blood of the sick dogs was 66% higher compared with the clinically healthy dogs (Table 1). At the same time, four out of the ten dogs were diagnosed with leukocytosis due to increase in band neutrophils.

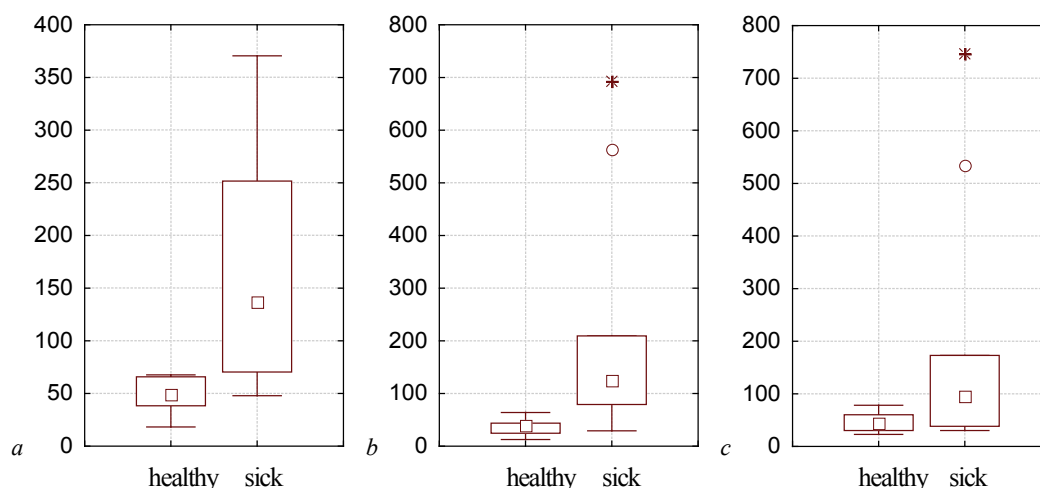
3.4 times higher ( $164.8 \pm 34.4 \text{ U/L}$ ;  $P < 0.01$ ) than in the clinically healthy animals ( $48.4 \pm 15.3 \text{ U/L}$ ). The activity of ALT in serum of the sick animals rose to  $215.9 \pm 71.4 \text{ U/L}$ , 5.9 times exceeding ( $P < 0.05$ ) the parameters in the clinically healthy animals ( $36.5 \pm 16.5 \text{ U/L}$ , Fig. 2c). At the same time, the AP activity in the blood of the patients was 4-fold higher ( $197.5 \pm 77.1 \text{ U/L}$ ) compared with the clinically healthy animals ( $45.0 \pm 18.7 \text{ U/L}$ , Fig. 2c).

The concentration of electrolytes in the blood serum varied little between the examined clinically healthy dogs and the sick dogs, but in most of the animals it was within the physiological fluctuations (Table 2). However, it has to be noted that the content of cations ( $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{++}$ ) in the patients tended to decline. In six of the ten dogs with signs of hepatic encephalopathy,  $\text{Na}^+$  ions in serum were below the lower threshold of the parameters of control dogs.

According to the ultrasound study of the liver, the clinically healthy animals had the typical anatomic configuration and distinct even margins. In the dogs suffering hepatic encephalopathy, we observed increase in the borders of the liver and smoothened and moderately expressed vascular pattern. Echogenicity of the parenchyma in them ranged moderately to significantly increased. The echostructure of the liver was characterized by non-homogeneity, some patients having significant granularity. No tumors were found in the liver and other organs of the abdominal cavity.



**Fig. 1.** Biochemical parameters of blood of the clinically healthy and sick dogs: *a* – ammonia content ( $\mu\text{mol/L}$ ) in plasm; *b* – content of bile acids ( $\mu\text{mol/L}$ ) in serum; *c* – content of bilirubin ( $\mu\text{mol/L}$ ) in serum; *d* – content of albumin ( $\text{g/L}$ ) in serum; abscissa axis indicates the groups of animals, ordinate axis shows the measurement units of the parameters; small square – median, upper and lower rectangle borders – 25% and 75% quartiles, vertical line – minimum and maximum values, circles – outliers;  $n = 10$



**Fig. 2.** Activity of enzymes in blood serum of the dogs (U/L): *a* – AST, *b* – ALT, *c* – AP abscissa axis indicates groups of animals, ordinate axis shows measurement units of the parameter; small square – median, upper and lower rectangle borders – 25% and 75% quartiles, vertical line – minimum and maximum values, circles – outliers;  $n = 10$

**Table 2**

Concentration of electrolytes (mmol/L) and pH in blood serum of the dogs ( $\bar{x} \pm \text{SE}$ ,  $n = 10$ )

Groups of dogs	$\text{Na}^+$	$\text{K}^+$	$\text{Cl}^-$	$\text{Ca}^{++}$	pH
Clinically healthy	$148.0 \pm 1.2$	$4.8 \pm 0.4$	$108.5 \pm 3.4$	$1.41 \pm 0.42$	$7.40 \pm 0.05$
Sick dogs	$145.8 \pm 3.1$	$4.6 \pm 0.4$	$107.8 \pm 1.7$	$1.35 \pm 0.03$	$7.39 \pm 0.06$

## Discussion

The data obtained as a result of the gathering anamneses, clinical studies, laboratory blood analyses, and ultrasound examination of the sick animals indicated concurrent lesions of the liver and the central nervous system. In particular, the sick animals were found to have symptoms that are typical for pathologies of the liver (pain and increase in the size of the liver) and the central nervous system (stupor, ataxia, spasms). The clinical signs of lesions of the liver and central nervous system suggest the development of hepatic encephalopathy (Tivers et al., 2014; Lidbury et al., 2016). Most often, hepatogenic encephalopathy has been diagnosed in Yorkshire Terriers, Miniature Schnauzers, Chihuahuas, Labrador Retrievers, Poodles, Pugs, Dachshunds, Cocker Spaniels, and Pomeranian dogs (Lidbury et al., 2015; Cheon et al., 2022). In our study of 10 sick dogs, the pathology was found in Yorkshire Terriers, two Maltese dogs, two Russian Toy dogs, one English Cocker Spaniel, and one mixed-breed dog. Therefore, our data are consistent with the literature sources.

The course of the disease in the dogs varied moderate to severe. This has been indicated by other researchers as well (Ferenci, 2017). Therefore,

it is important to gather a comprehensive anamnesis and to thoroughly examine the patient clinically in order to assess the complexity of the pathological process and its duration (Xenoulis et al., 2015; Jawaro et al., 2016). During insignificant changes in the clinical status of the patients, when the symptoms of damage to the central nervous system are not clearly apparent, important data are provided by laboratory blood assays. Blood analysis can reveal the severity of liver malfunctioning, in particular, in neutralizing toxins that cause harm to the brain (Hadjihambi et al., 2019; Nardelli et al., 2023). Ammonia is an endotoxin, which inflicts the most harmful damage on the central nervous system (Webster, 2017; Kawaguchi et al., 2019; European Association, 2022). We determined that ammonia analysis is a highly informative test of concurrent damages to the liver and central nervous system. Hyperammonemia was closely associated with the severity of the clinical course of hepatic encephalopathy, and also disorders in the main functions of the liver. The content of ammonia in the blood is high in cases of severe hepatitis, cirrhosis, or necrosis of the liver, indicating the presence of congenital or acquired portosystemic shunts (Chapman et al., 2013; Seller et al., 2022; Farhoodimoghdam et al., 2024).

Besides ammonia, important parameters for diagnosing liver lesions are the contents of albumin, bile acids, bilirubin, urea, and also the activities of alanine aminotransferase, aspartate aminotransferase, and alkaline phosphatase in the blood (Simonov & Vlizlo, 2015; Lawrence & Steiner, 2017; Zelenina et al., 2022). Therefore, we studied the blood serum of the sick dogs, finding decreased content of albumin ( $P < 0.05$ ), which is a sign of malfunctioning of the most important activity of the liver – synthesis of protein. At the same time, the sick animals were observed to have cholelithiasis and hyperbilirubinemia. This is explained by the fact that damage to the hepatic cells leads to disorder in the transport of bile acids and bilirubin from hepatocytes to the bile ducts, causing enhanced inflow of those metabolites into the blood (Selgas et al., 2014; Lester et al., 2016; Vlizlo et al., 2023). It is also possible that significant increase in the concentrations of bile acids and bilirubin in the blood of patients can also cause toxic impact on the nervous system.

It was reported that the contributing factors of the development of hepatic encephalopathy can be inflammatory cytokines (Odeh, 2007; Kilpatrick et al., 2014), hyponatremia, and hypokalemia (Tivers et al., 2014; Lidbury et al., 2016). This was corroborated by our data. In particular, in the blood serum of the sick dogs with severe pathology, the concentrations of  $\text{Na}^+$  and  $\text{K}^+$  ions were low. Hyponatremia causes brain edema, and hypokalemia enhances the renal ammoniogenesis and decreases the renal excretion of ammonia (Gow, 2017), thereby exacerbating the pathology. At the same time, in half of the patients, we found leukocytosis due to increase in neutrophils, indicating inflammatory processes in the body. Those animals also had lymphocytopenia, evidencing weakened protective powers of the body (Elhiblu et al., 2015).

Of the hematological parameters, we should note reduced content of hemoglobin in the sick dogs. In two of the ten, we found erythrocytopenia and decline in hematocrit value. Such changes in blood can indicate the onset of anemia in certain animals. This was observed during the development of pathological processes in the liver (Webster et al., 2019) and could be associated with hemorrhage in the gastrointestinal tract in hepatic encephalopathy patients (Lidbury et al., 2015).

Significant damage to the hepatic cells was suggested by stable increase in the activity of aminotransferase (AST, ALT) and AP in the blood serum of the sick dogs. This was also pointed out by other scientists (Alvarez & Whittemore, 2009; Vallarino et al., 2020). The first cells to be damaged are hepatocytes that come into contact with blood inflowing from the gastrointestinal tract, which contains a large amount of ammonia and other endotoxins (Jalan et al., 2006; Jawaro et al., 2016; Krishnarao & Gordon, 2020). At the same time, increase in the echotexture of parenchyma and disorders in hemodynamics of the organ, concurring with high activity of the indicator enzymes, confirm the damage to the hepatic cells (Assawarachan et al., 2019).

## Conclusions

The liver disease in the dogs manifested in disorders of the neutralizing function of hepatocytes with accumulation of ammonia in the blood, which was harmful to the central nervous system, causing the development of hepatic encephalopathy. During this pathology, we observed the typical symptoms of liver lesion (pain and increase in the size of the liver), changes in the structure, and disruptions of the main functions of hepatocytes (neutralizing, protein-synthesizing, bile-forming, and bile-excreting), and also the typical symptoms of damage to the brain (stupor, ataxia, spasms). In cases of severe hepatic encephalopathy, the blood of the sick dogs had decreased ions of sodium, leukocytosis developed, and also neutrophilia, lymphocytopenia, and signs of anemia. In the future, it is important to determine the role of portosystemic shunts in the development of hepatic encephalopathy in dogs.

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## References

Alvarez, L., & Whittemore, J. (2009). Liver enzyme elevations in dogs: Diagnostic approach. *Compendium: Continuing Education for Veterinarians*, 31(9), 416–418.

- Assawarachan, S. N., Chuchalempom, P., Maneesaay, P., & Thengchaisri, N. (2019). Evaluation of hepatobiliary ultrasound scores in healthy dogs and dogs with liver diseases. *Veterinary World*, 12(8), 1266–1272.
- Assawarachan, S. N., Maneesaay, P., & Thengchaisri, N. A. (2020). Descriptive study of the histopathologic and biochemical liver test abnormalities in dogs with liver disease in Thailand. *Canadian Journal of Veterinary Research*, 84(3), 217–224.
- Assawarachan, S. N., Ongvisespaibool, T., Hakhen, B., Chuchalempom, P., Maneesaay, P., & Thengchaisri, N. (2023). Predictive factors for two-year survival in dogs with hepatobiliary diseases: Importance of clinical and laboratory monitoring. *Animals*, 13(16), 2677.
- Bellafante, D., Gioia, S., Faccioli, J., Riggio, O., Ridola, L., & Nardelli, S. (2024). The management of hepatic encephalopathy from ward to domiciliary care: Current evidence and gray areas. *Journal of Clinical Medicine*, 13(1), 166.
- Besa, C., Cruz, J. P., Huete, A., & Cruz, F. (2012). Portal biliopathy: A multitechnique imaging approach. *Abdominal Imaging*, 37, 83–90.
- Caporali, E. H. G., Phillips, H., Underwood, L., & Selmic, L. E. (2015). Risk factors for urolithiasis in dogs with congenital extrahepatic portosystemic shunts: 95 cases (1999–2013). *Journal of the American Veterinary Medical Association*, 246, 530–536.
- Chapman, S. E., & Hostutler, R. A. (2013). A laboratory diagnostic approach to hepatobiliary disease in small animals. *Veterinary Clinics of North America: Small Animal Practice*, 43, 1209–1225.
- Cheon, S. Y., Jo, D., Kim, Y. K., & Song, J. (2022). Long noncoding RNAs regulate hyperammonemia-induced neuronal damage in hepatic encephalopathy. *Oxidative Medicine and Cellular Longevity*, 2022, 7628522.
- Elhiblu, M. A., Dua, K., Mohindroo, J., Mahajan, S. K., Sood, N. K., & Dhaliwal, P. S. (2015). Clinico-hemato-biochemical profile of dogs with liver cirrhosis. *Veterinary World*, 8, 487–491.
- European Association for the Study of the Liver (2022). EASL Clinical Practice Guidelines on the management of hepatic encephalopathy. *Journal of Hepatology*, 77(3), 807–824.
- Farhoodmoghadam, M., Reagan, K. L., & Zwingerberger, A. L. (2024). Diagnosis and classification of portosystemic shunts: A machine learning retrospective case-control study. *Frontiers in Veterinary Science*, 11, 1291318.
- Ferenci, P. (2017). Hepatic encephalopathy. *Gastroenterology Report*, 5(2), 138–147.
- Gluud, L. L., Vilstrup, H., & Morgan, M. Y. (2016). Nonabsorbable disaccharides for hepatic encephalopathy: A systematic review and meta-analysis. *Hepatology*, 64, 908–922.
- Gow, A. G. (2017). Hepatic encephalopathy. *Veterinary Clinics of North America: Small Animal Practice*, 47(3), 585–599.
- Hadjihambi, A., Harrison, I. F., Costas-Rodríguez, M., Vanhaecke, F., Arias, N., Gallego-Durán, R., Mastitskaya, S., Hosford, P. S., Olde Damink, S. W. M., Davies, N., Habtesion, A., Lythgoe, M. F., Gourine, A. V., & Jalan, R. (2019). Impaired brain lymphatic flow in experimental hepatic encephalopathy. *Journal of Hepatology*, 70(1), 40–49.
- Jalan, R., Shawcross, D., & Davies, N. (2003). The molecular pathogenesis of hepatic encephalopathy. *The International Journal of Biochemistry and Cell Biology*, 35(8), 1175–1181.
- Jawaro, T., Yang, A., Dixit, D., & Bridgeman, M. B. (2016). Management of hepatic encephalopathy: A primer. *Annals of Pharmacotherapy*, 50(7), 569–577.
- Kabaria, S., Dalal, I., Gupta, K., Bhurwal, A., Minacapelli, C. D., Catalano, C., & Rustgi, V. (2021). Hepatic encephalopathy. *Hepatology*, 9(1), 89–97.
- Kashliak, N. O., & Vlizlo, V. V. (2023). Symptomy, biokhimichni pokaznyky ta zahal'nyj analiz krovi za hepatopatiji u sobak [Symptoms, biochemical indicators and general blood analysis for hepatopathy in dogs]. *Scientific Messenger of Lviv National University of Veterinary Medicine and Biotechnologies. Series: Veterinary Sciences*, 112, 193–200 (in Ukrainian).
- Kawaguchi, T., Suzuki, F., Imamura, M., Murashima, N., Yanase, M., Mine, T., Fujisawa, M., Sato, I., Yoshiji, H., Okita, K., & Suzuki, K. (2019). Rifaximin-altered gut microbiota components associated with liver/neuropsychological functions in patients with hepatic encephalopathy: An exploratory data analysis of phase II/III clinical trials. *Hepatology Research*, 49(4), 404–418.
- Kilpatrick, S., Gow, A. G., Foale, R. D., Tappin, S. W., Caruthers, H., Reed, N., Yool, D. A., Woods, S., Marques, A. I., Jalan, R., Mellanby, R. J. (2014). Plasma cytokine concentrations in dogs with a congenital portosystemic shunt. *The Veterinary Journal*, 200(1), 197–199.
- Konstantinidis, A. O., Patsikas, M. N., Papazoglou, L. G., & Adamama-Moraitou, K. K. (2023). Congenital portosystemic shunts in dogs and cats: Classification, pathophysiology, clinical presentation and diagnosis. *Veterinary Sciences*, 10(2), 160.
- Kraun, M. B., Nelson, L. L., Hauptman, J. G., & Nelson, N. C. (2014). Analysis of the relationship of extrahepatic portosystemic shunt morphology with clinical variables in dogs: 53 cases (2009–2012). *Journal of the American Veterinary Medical Association*, 245(5), 540–549.
- Krishnarao, A., & Gordon, F. D. (2020). Prognosis of hepatic encephalopathy. *Clinics in Liver Disease*, 24(2), 219–229.



- Lawrence, Y. A., & Steiner, J. M. (2017). Laboratory evaluation of the liver. *Veterinary Clinics of North America: Small Animal Practice*, 47, 539–553.
- Lester, C., Cooper, J., Peters, R. M., & Webster, C. R. L. (2016). Retrospective evaluation of acute liver failure in dogs (1995–2012): 49 cases. *Veterinary Emergency and Critical Care Society*, 26(4), 559–567.
- Levitt, D. G., & Levitt, M. D. (2018). A model of blood-ammonia homeostasis based on a quantitative analysis of nitrogen metabolism in the multiple organs involved in the production, catabolism, and excretion of ammonia in humans. *Clinical and Experimental Gastroenterology*, 11, 193–215.
- Lidbury, J. A., Cook, A. K., & Steiner, J. M. (2016). Hepatic encephalopathy in dogs and cats. *Veterinary Emergency and Critical Care Society*, 26(4), 471–487.
- Lidbury, J. A., Ivanek, R., Suchodolski, J. S., & Steiner, J. M. (2015). Putative precipitating factors for hepatic encephalopathy in dogs: 118 cases (1991–2014). *Journal of the American Veterinary Medical Association*, 247(2), 176–183.
- Lima, L. C. D., Miranda, A. S., Ferreira, R. N., Rachid, M. A., & Simões e Silva, A. C. (2019). Hepatic encephalopathy: Lessons from preclinical studies. *World Journal of Hepatology*, 11(2), 173–185.
- Lynch, A. (2023). Hepatic encephalopathy. *Small Animal Critical Care Medicine*, 506–509.
- Mullins, R. A., Carrera, A. E., Anderson, D. M., & Billet, J.-P. (2022). Postattenuation neurologic signs after surgical attenuation of congenital portosystemic shunts in dogs: A review. *Veterinary Surgery*, 51(1), 23–33.
- Nardelli, S., Bellafante, D., Ridola, L., Faccioli, J., Riggio, O., & Gioia, S. (2023). Prevention of post-tips hepatic encephalopathy: The search of the ideal candidate. *Metabolic Brain Disease*, 38(5), 1729–1736.
- Odeh, M. (2007). Pathogenesis of hepatic encephalopathy: The tumour necrosis factor-alpha theory. *European Journal of Clinical Investigation*, 37(4), 291–304.
- Romero-Gómez, M., Montagnese, S., & Jalan, R. (2015). Hepatic encephalopathy in patients with acute decompensation of cirrhosis and acute-on-chronic liver failure. *Journal of Hepatology*, 62(2), 437–447.
- Rose, C. F., Amodio, P., Bajaj, J. S., Dhiman, R. K., Montagnese, S., Taylor-Robinson, S. D., Vilstrup, H., & Jalan, R. (2020). Hepatic encephalopathy: Novel insights into classification, pathophysiology and therapy. *Journal of Hepatology*, 73(6), 1526–1547.
- Rothuizen, J. (2009). Important clinical syndromes associated with liver disease. *Veterinary Clinics of North America: Small Animal Practice*, 39(3), 419–437.
- Selgas, A. G., Bexfield, N., Sease, T. J., Holmes, M. A., & Watson, P. (2014). Total serum bilirubin as a negative prognostic factor in idiopathic canine chronic hepatitis. *Journal of Veterinary Diagnostic Investigation*, 26(2), 246–251.
- Seller, S., Weisse, C., & Fischetti, A. J. (2022). Intrahepatic venous collaterals in dogs with congenital intrahepatic portosystemic shunts are associated with focal shunt or hepatic vein narrowing. *Veterinary Radiology and Ultrasound*, 63(1), 64–72.
- Simonov, M., & Vlizlo, V. (2015). Some blood markers of the functional state of liver in dairy cows with clinical ketosis. *Bulgarian Journal of Veterinary Medicine*, 18(1), 74–82.
- Swaminathan, M., Ellul, M. A., & Cross, T. J. (2018). Hepatic encephalopathy: Current challenges and future prospects. *Hepatic Medicine: Evidence and Research*, 10, 1–11.
- Tivers, M. S., Handel, I., Gow, A. G., Lipscomb, V. J., Jalan, R., & Mellanby, R. J. (2014). Hyperammonemia and systemic inflammatory response syndrome predicts presence of hepatic encephalopathy in dogs with congenital portosystemic shunts. *PLoS One*, 9(1), e82303.
- Vallarino, N., Pil, S., Devriendt, N., Or, M., Vandermeulen, E., Serrano, G., Paepe, D., Bosmans, T., & de Rooster, H. (2020). Diagnostic value of blood variables following attenuation of congenital extrahepatic portosystemic shunt in dogs. *Veterinary Record*, 187(7), e48.
- Vlizlo, V. V. (1999). Hepatozerebralnyj syndrom u velykoyi rohatoyi khudoby [Hepatocerebral syndrome in cattle]. *Visnyk Ahramoyi Nauky*, 8, 29–33 (in Ukrainian).
- Vlizlo, V., Prystupa, O., Slivinska, L., Gutyj, B., Maksymovych, I., Shcherbatyy, A., Lychuk, M., Partyka, U., Chernushkin, B., Rusyn, V., Leno, M., & Leskiv, K. (2023). Treatment of animals with fatty liver disease using a drug based on the seeds of *Silybum marianum*. *Regulatory Mechanisms in Biosystems*, 14(3), 424–431.
- Wang, A. J., Peng, A. P., Li, B. M., Gan, N., Pei, L., Zheng, X. L., Hong, J. B., Xiao, H. Y., Zhong, J. W., & Zhu, X. (2017). Natural history of covert hepatic encephalopathy: An observational study of 366 cirrhotic patients. *World Journal of Gastroenterology*, 23(34), 6321–6329.
- Webster, C. R. L. (2017). Hemostatic disorders associated with hepatobiliary disease. *Veterinary Clinics of North America: Small Animal Practice*, 47, 601–615.
- Webster, C. R. L., Center, S. A., Cullen, J. M., Penninck, D. G., Richter, K. P., Twedt, D. C., & Watson, P. J. (2019). ACVIM consensus statement on the diagnosis and treatment of chronic hepatitis in dogs. *Journal of Veterinary Internal Medicine*, 33(3), 1173–1200.
- Xenoulis, P. G., & Steiner, J. M. (2015). Canine hyperlipidaemia. *Journal of Small Animal Practice*, 56(10), 595–605.
- Zelenina, O., Vlizlo, V., Kozak, M., Ostapiv, D., Samaryk, V., Dron, I., Stetsko, T., Skrypka, M., Tomchuk, V., Danchuk, O., & Levchenko, A. (2022). Antimicrobial activity of the PEGylated antibiotic enrofloxacin and its functional and structural effect on the liver in rats. *Journal of Applied Pharmaceutical Science*, 12(6), 68–75.